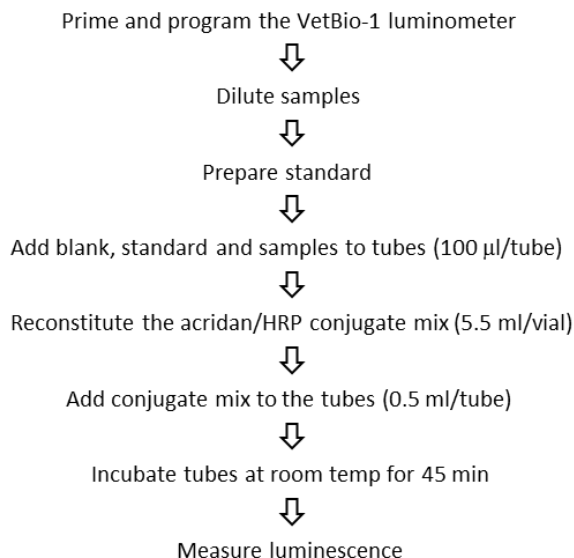


INTRODUCTION

Serum amyloid A (SAA) is a positive acute phase protein that is expressed in liver and circulates in blood. In cats it can increase >1000-fold during inflammation and infection.

PRINCIPLE OF THE ASSAY

The cat SAA assay uses SPARCL™¹ technology. Two SAA antibodies are used. One is conjugated to horseradish peroxidase (HRP), the other to acridan, a chemiluminescent substrate. When HRP and acridan conjugates bind to SAA they are brought into proximity. Upon addition of hydrogen peroxide, HRP catalyzes oxidation of proximal acridan molecules causing a flash of luminescence that is proportional to SAA concentration. Diluted serum or plasma samples and standards are dispensed into test tubes and mixed with 0.5 ml of acridan and HRP conjugates. After 45 minutes, tubes are placed in the VetBio-1 luminometer. Luminescence is measured after injection of a background reducer, that eliminates nonspecific luminescence, followed by trigger-solution containing hydrogen peroxide. The concentration of SAA is automatically determined from the ratio of sample luminescence to that of the 25 ng/ml standard. The assay sequence is illustrated below.



KIT COMPONENTS

Materials provided with the kit:

- Acridan & HRP conjugates, 5 vials **Store at -20°C**
- SAA stock, 1 vial **Store at -20°C**
- Diluent: CSD50-1, 2 x 50 ml
- Background reducer: BR9-1, 9 ml
- Trigger solution: TS12-1, 12 ml
- 15 ml centrifuge tubes, 2
- 5 ml centrifuge tubes, 2

Materials required but not provided:

- VetBio-1 luminometer
- Precision pipettors and tips
- 12 x 75 mm borosilicate tubes²
- 12 x 75 mm test tube racks

STORAGE

Store the conjugate and SAA stock vials at or below -20°C. The remainder of the kit should be stored at 4°C.

GENERAL INSTRUCTIONS

1. Please take the time to completely read all instructions before starting your assay. Contact us if you need clarification.
2. All reagents used in the assay should be allowed to reach room temperature before use.

LUMINOMETER SETUP

1. Turn the VetBio-1 luminometer on.
2. Place the tubing from injector 1 into the tube containing background reducer. Tube holders are positioned adjacent to the injectors.
3. Place the tube from injector 2 into the tube containing trigger solution.
4. From the Protocol manager on the keypad select "Prime & Wash", "Prime", then "Start".
5. Open the luminometer drawer and insert an empty 12 x 75 mm tube.
6. Close the drawer and click "Start" again.
7. Wait for priming to complete, open the drawer and discard the tube.
8. From the Protocol manager select "Measure" followed by "Cat SAA".

¹ SPARCL technology, using acridan- and HRP-conjugated antibodies, was developed by and is licensed from Lumigen Corp.

² Only cylindrical 12 x 75 mm borosilicate glass tubes can be used. Do not use polystyrene or polycarbonate tubes.

9. Select "Start".
10. The experiment setup screen will be displayed, allowing you to enter experiment name, comments, and the number of samples. After entering the information select "Start".
11. Enter the sample IDs and dilution factor(s), if different than the default dilution factor.
12. The luminometer is now ready for use.
13. Press "Start" when you are ready to measure luminescence.

SAMPLE PREP

This assay was designed for measurement of SAA in serum. We recommend testing at a 100-fold dilution. Prepare a 100-fold dilution of each sample as follows.

1. Dispense 0.99 ml of CSD50-1 into a glass test tube.
2. Add 10 μ l of serum and mix gently.
3. Use the samples within 10 minutes.

STANDARD PREP

Prepare the 25 ng/ml standard by diluting 4.0 μ l of the SAA stock with 3.996 ml of CSD50-1 diluent.

CONJUGATE MIX PREP

1. The acridan and HRP conjugate mix should be prepared just before use (step 5 in the Procedure section).
2. Tap the vial to ensure that the contents are at the bottom of the vial before carefully removing the stopper.
3. Add 5.5 ml of diluent CSD50-1 to the vial. Insert the stopper and mix gently by inversion several times.
4. Each vial of reconstituted conjugate mix provides enough reagent to measure 0 and 25 ng/ml standards and up to eight samples.³

PROCEDURE

1. Determine the number of 12 x 75 mm borosilicate glass tubes required for the assay. Ensure that all tubes fit easily in the VetBio-1 sample holder.
2. Pipet 100 μ l of diluent into assay tube one. This serves as the zero standard.
3. Pipet 100 μ l of the 25 ng/ml SAA standard into tube two.
4. Pipet 100 μ l aliquots of the diluted samples into tubes 3, 4, 5... as defined by your assay format.
5. Add 0.50 ml of freshly prepared conjugate mix to each tube and mix gently. A vortex mixer may be used if available but is not necessary.
6. Incubate the mixtures at room temperature.
7. After 45 minutes insert tube 1 into the sample holder of the luminometer and close the drawer. The luminometer automatically injects 50 μ l of background reducer and 100 μ l of trigger solution, then measures luminescence (RLU/s).
8. Once the RLU/s value is recorded on the screen open the drawer and discard the tube.
9. Determine luminescence for the remaining tubes.
10. SAA concentrations are automatically calculated.
11. After measurement of the last sample, select "End".
12. Results may be exported via a USB stick in Excel, HTML, or pdf format.

LUMINOMETER MAINTENANCE

The luminometer injectors must be cleaned with distilled or deionized water at the end of each day of use to avoid clogging of the injector ports.

1. From the Protocol manager screen select "Prime & Wash".
2. Select "Backprime" to return unused reagents to the respective tubes.
3. If future use of the reagents is intended store the sealed tubes in a refrigerator.
4. Place the tubing from injectors 1 and 2 into separate 15 ml centrifuge tubes containing distilled or deionized water.
5. Select "Wash" from the "Prime & Wash" screen. Press "Start", insert an empty tube into the luminometer, close the drawer and press "Start".
6. Discard the tube.
7. Leave the injector tubing immersed in water.
8. Switch the luminometer off. It should not be left on overnight.

REPLICATES

The VetBio-1 luminometer protocols are configured to run singlets of the blank, standard and samples. At the discretion of the user it can be configured to run replicates.

1. From the protocol manager select "Measure" and the program you would like to modify.
2. Select "New" and "Copy Protocol".
3. Increase the Replicates as desired.
4. Create a new protocol name.
5. Select "Protocols" and save the new protocol.

³ If using multiple vials of conjugates to measure more than eight samples, combine the reconstituted contents of all vials and mix briefly before dispensing 0.5 ml aliquots into the reaction tubes. Larger volumes of background reducer and trigger solution must also be used.

ASSAY PERFORMANCE

Typical data.

Results are shown below from an assay in which serum samples from three cats were tested at a 10000-fold dilution, as singlets.



**Veterinary
Biomarkers, Inc.**

Cat SAA Exp#002

18.12.2019 22:28:58

Single Assay	Start by :	DOOR	Blank :	ON
Measurement	Delay [s] :	2.600	Time [s] :	0.800
Injector 1	Delay [s] :	1.000	Volume [μL] :	50
Injector 2	Delay [s] :	2.000	Volume [μL] :	100

Experiment comment: Serum test

Sample	Dilution	Rep	RLU/s	Conc [μg/mL]
Blank			2,325	0
Standard			545,518	0.025
1	10000		114,059	51.425
2	10000		4,020	0.78011
3	10000		17,185	6.8392

Reproducibility. Three serum samples with different SAA levels were tested in triplicate (Table 1) or as singlets (Table 2) in three separate assays at dilutions of 100, 1000, or 10000-fold. Levels of SAA are reported as μg/ml (mean ± SD, in Table 1).

Table 1.

Sample	Dilution	Assay 1	Assay 2	Assay 3
1	10000	53.7±5.4	72.3±15.4	67.6±2.4
2	100	0.35±0.04	0.45±0.14	0.38±0.01
3	1000	4.82±0.02	5.99±1.12	5.57±0.17

Table 2.

Sample	Dilution	Assay 1	Assay 2	Assay 3
1	10000	55.5	56.4	66.8
2	100	0.32	0.61	0.37
3	1000	4.84	5.50	5.37

FREQUENTLY ASKED QUESTIONS

- Can I dilute the standard and samples, then use them later?
 - No. Once diluted in the assay buffer, SAA has limited stability; after one hour, recovered values decrease by approximately 10%. Diluted standard and samples should be used within 10-minutes.
- Can I reconstitute the acridan/HRP conjugate reagent and use it later?
 - No. When reconstituted, the conjugate mix has limited stability. Luminescence signals decrease by approximately 25% after one hour. The reconstituted conjugate mix should be used within 10-minutes.
- How stable is SAA in serum?
 - We find that SAA is quite stable in serum, samples can be stored in tightly sealed vials for at least one day at 4°C and at least one year at -20°C.
- How stable is the SAA stock?
 - The SAA stock is stable for at least 10 days at room temperature and at least one year at -20°C.
- How stable are the Background Reducer and Trigger Solution?
 - Both are stable at room temperature for several days. However, after use they should be stored in capped vials in the refrigerator. Be careful not to switch caps.

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